

## The probiotic *Saccharomyces boulardii* upgrades intestinal digestive functions by several mechanisms

Jean-Paul Buts

Laboratory of Pediatric Gastroenterology and Nutrition, Faculty of Medicine, Université Catholique de Louvain, B-1200, Brussels, Belgium.

**Key words:** Microvillous enzymes, *S. boulardii*, probiotic, polyamines, protein phosphatase, secretory IgA, SGLT-1, disaccharidases.

**Abbreviations:** BBM: Brush Border Membrane; *S. boulardii*: *Saccharomyces boulardii*; MAP kinase: Mitogen activating kinase; ERK<sub>1</sub>–ERK<sub>2</sub>: Extra membrane signal regulating kinase 1,2; SGLT<sub>1</sub>: Sodium glucose cotransporter –1.

To the Editor,

*Saccharomyces boulardii* (*S. boulardii*) is a probiotic yeast largely prescribed in many countries over the world. This probiotic has shown to exhibit therapeutic properties in acute and chronic enterocolopathies, irritable bowel syndrome, antibiotic – associated – diarrhoea and enterotoxigenic *Clostridium difficile* infections (1). However, the mechanisms of its effects remain largely unknown. The aim of this letter is to propose three complementary mechanisms explaining its trophic effects. In human volunteers (2) and growing rats (2), oral administration of lyophilized preparations of *S. boulardii* produces trophic intestinal effects including increases in the specific and total activities of sucrase and maltase-glucoamylase in epithelial cells and in endoluminal fluid, enhanced secretion of s-IgA in intestinal fluid (3) and enhanced synthesis of the secretory component (receptor for polymeric immunoglobulins) in crypt cells (3). Also, rats treated with lyophilized preparations of *S. boulardii* show a marked stimulation of the sodium – dependent D-glucose uptake with a corresponding increase in the sodium D-glucose co-transporter 1 (SGLT<sub>1</sub>) (4). Recently we demonstrated that yeast cells can release in the endoluminal compartment a sucrase proteinase (2, 6), a leucine aminopeptidase acting as a zinc-binding metalloprotease (6) and a novel protein phosphatase that inhibits *Escherichia coli* endotoxin by dephosphorylation (7). Stimulation of microvillous enzymes by *S. boulardii* was confirmed in human enterocytes by an in situ histochemistry method (8). Since *S. boulardii* secretes a great amount of sucrase, totalizing more than 8000 units per g of lyophilized preparation (2), yeast cells are used to treat children with congenital sucrase-isomaltase deficiency (9). In addition, lyophilized *S. boulardii* is recognized by OMS authorities as an effective treatment for *Clostridium difficile* overgrowth (1).

To complete the digestion of sugars, *S. boulardii* delivers an  $\alpha,\alpha$ -trehalase in the intestinal lumen of rats which could be efficient to treat trehalose intolerance (10). Expressed per gram of powder,  $\alpha,\alpha$ -trehalase from *S. boulardii* delivered in vitro an activity 175 times higher than the activity of human trehalase expressed per gram of intestinal mucosa (10).

Like as with sucrase, the luminal secretion of an  $\alpha$ -amylase (11) acting on starch  $\alpha$ , 1-4 bounds and of a maltase-glucoamylase (2,11) acting on the last glucosyl-unit of each amylopectin chain could implicate oral treatment of *S. boulardii* in the digestion and metabolic degradation of starch.

Beside the secretion of enzymes and of nutrient carriers in the intestinal lumen, the stimulation of brush border membrane enzymes and of carriers is at least in part mediated by the endoluminal release of polyamines (12) mainly spermine and spermidine. Lyophilised preparation of yeasts contain significant quantities of polyamines, totalling 679 nanomoles/100mg of lyophilised preparation mainly spermidine (55%) and spermine (43%) with negligible amounts of putrescine (1.4%). In theory, such amounts of polyamines could influence intestinal expression of brush border membrane glycoproteins (hydrolases, proteases and transport molecules). In practice, a marked stimulation of disaccharidases and aminopeptidase activities and of the secretion of secretory IgA has been observed in the small intestine of young unweaned rats in response to oral ingestion of spermine and spermidine, equivalent to 1000 nanomoles/day of purified polyamines (12). When infant rats were given an amount of spermine (500 nanomoles/day) equivalent to the polyamine content of the lyophilised preparation of yeast cells (679 nanomoles/100 mg) similar enzymatic responses were observed, including significant increases in the specific and total activities of sucrase ( $\times 2,5$ ) and maltase

Correspondence to: Prof. Jean-Paul Buts, M.D., Ph.D., Université Catholique de Louvain, Laboratory of Pediatric Gastroenterology and Nutrition, Tour Pasteur +3, 53, avenue Mounier, B-1200 Bruxelles, Belgium.  
E-mail: buts@gype.ucl.ac.be

Work supported by grants from the Laboratory Biocodex, Gentilly, France.

Submission date: 18/02/2009

Acceptance date: 27/04/2009

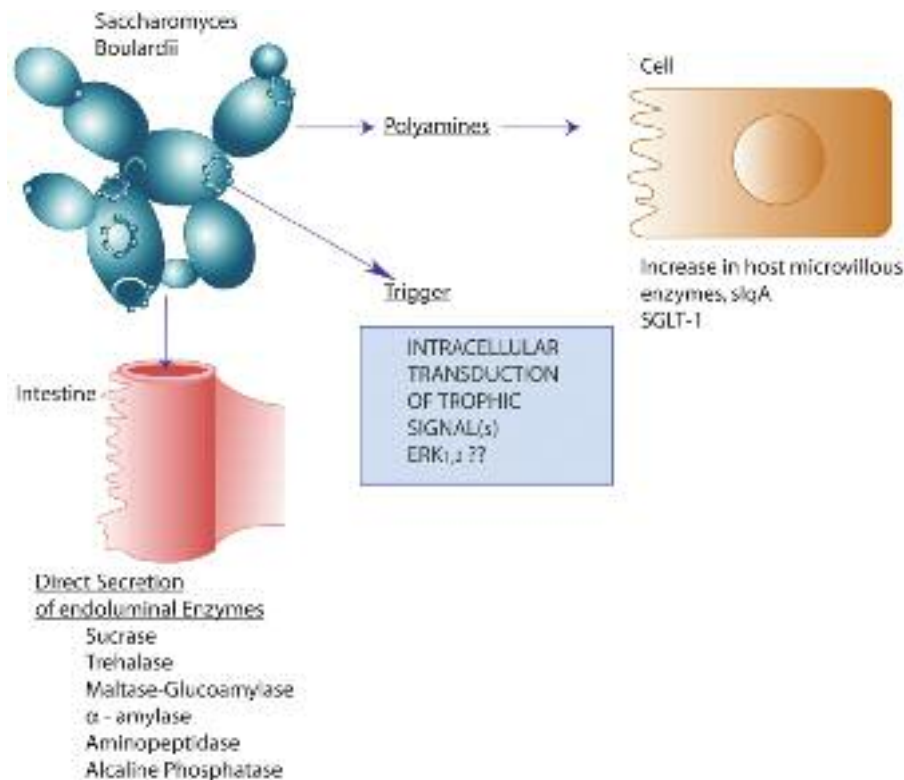


Fig. 1. — The Figure represents the three mechanisms of action of the probiotic *Saccharomyces boulardii*

(+ 24%). In response to 1000 nanomoles of spermine, enzyme stimulation was proportionally greater with increases in sucrase ( $\times 4,6$ ) and maltase (+ 70%) activities. Similarly, weaned rats treated with either *S. boulardii* or equivalent quantities of spermine (500 nanomoles) presented parallel, significant increases in specific sucrase (157%) and maltase (+ 47,5%) activities. Therefore, oral administration of 100 mg lyophilised *S. boulardii* containing 679 nanomoles of polyamines to breast fed sucklings reproduced similar changes in microvillous enzymes than the administration of 500 nanomoles of spermine. Regarding the stimulation of microvillous enzymes, the oral administration of spermine is a dose – dependent phenomenon. It is more sensitive for sucrase than for other microvillous enzymes (lactase, aminopeptidase, maltase-glucoamylase) and becomes detectable at doses of spermine as low as 250 nanomoles/day. After absorption, polyamines act at the level of DNA synthesis, mRNA expression and protein production. Lastly, recent experiments indicate that *S. boulardii* inhibits the signal transduction triggered by *E. coli* on in vitro intestinal cell lines, essentially interleukin 8 and NF- $\kappa$ B (13).

Up to now, the precise mechanism by which yeast cells triggers signals transduced in epithelial cells remains unknown. However, the ERK<sub>1</sub> and ERK<sub>2</sub> MAP kinases could be involved because these messengers are activated at the epithelial cell membrane by trophic signals and transduce them into the cell to the nucleus (14,

15). Furthermore, the disaccharidase activities are inhibited by specific ERK inhibitors (14). Further studies are warranted to determine the nature of the signal(s) transduced into epithelial cells.

**In conclusion :** oral administration of lyophilised *S. boulardii* upgrades intestinal functions mainly by three mechanisms : 1) the endoluminal secretion of enzyme proteins by the yeast itself ; 2) the endoluminal secretion of polyamines (spermine and spermidine) that after absorption enhances the synthesis of BBM proteins, enzymes and carriers ; and 3) the possible intracellular activation of messengers which transduce trophic signals from the apical membrane to the nucleus.

### Acknowledgements

The authors are grateful to P. Bernasconi and B. Hublot from Biocodex for their helpful comments on the manuscript.

### References

1. BUTS J.P., BERNASCONI P. *Saccharomyces boulardii* : basic science and clinical applications in gastroenterology. *Gastroenterol. Clin. North Am.*, 2005, **34** : 515-532.
2. BUTS JP., BERNASCONI P., VAN CRAYNEST MP., MALDAGUE P., DE MEYER R. Response of human and rat small intestinal mucosa to oral administration of *Saccharomyces boulardii*. *Pediatr. Res.*, 1986, **20** : 192-196.
3. BUTS JP., BERNASCONI P., VAERMAN JP., DIVE C. Stimulation of secretory IgA and secretory component of immunoglobulins in small intestine of

- rats treated with *Saccharomyces boulardii*. *Dig. Dis. Sci.*, 1990, **35** : 251-256.
4. BUTS J.P., DE KEYSER N., MARANDI S., HERMANS D., SOKAL E.M., CHAE Y.H.E., LAMBOTTE L., CHANTEUX H., TULKENS P.M. *Saccharomyces boulardii* upgrades cellular adaptation after proximal enterectomy in rats. *Gut*, 1999, **45** : 89-96.
  5. BUTS JP., DE KEYSER N., DE RAEDEMAEKER L. *Saccharomyces boulardii* enhances rat intestinal enzyme expression by endoluminal release of polyamines. *Pediatr. Res.*, 1994, **36** : 522-527.
  6. BUTS J.P., DE KEYSER N., STILMANT C., SOKAL E.M., MARANDI S. *Saccharomyces boulardii* enhances N-terminal peptide hydrolysis in suckling rat small intestine by endoluminal release of a zinc-binding metalloprotease. *Pediatr. Res.*, 2002, **51** : 528-534.
  7. BUTS J.P., DEKEYSER N., STILMANT C., DELEM F., SMETS F., SOKAL E.M. *Saccharomyces boulardii* produces in rat small intestine a novel protein phosphatase that inhibits *Escherichia coli* endotoxin by dephosphorylation. *Pediatr. Res.*, 2006, **60** : 24-29.
  8. JAHN H.U., ULRICH R., SCHNEIDER T. Immunology and topical effects of *S. boulardii* on the small intestine in healthy human volunteers. *Digestion*, 1996, **57** : 95-104.
  9. HARMS H.K., BERTELE-HARMS R.M., BRUER-KLEIS D. Enzyme-substitution therapy with the yeast *Saccharomyces cerevisiae* in congenital sucrase-isomaltase deficiency. *N. Engl. J. Med.*, 1987, **316** : 1306-1309.
  10. BUTS JP., STILMANT C., BERNASCONI P., NEIRINCK C., DE KEYSER N. Characterization of alpha,alpha-trehalase released in the intestinal lumen by the probiotic *Saccharomyces boulardii*. *Scand. J. Gastroenterol.*, 2008, **43** : 1489-1496.
  11. EKSTEEN J.M., VAN RENSBURG P., CORDERO-OTERO R.R., PRETORIUS I.S. Starch fermentation by recombinant *Saccharomyces cerevisiae* strains expressing the  $\alpha$ -amylase and glucoamylase genes from *Lipomyces kononenkoae* and *Saccharomycopsis fibuligera*. *Biotechnol. Bioeng.*, 2003, **84** : 639-646.
  12. BUTS JP., DE KEYSER N., KOLANOWSKI J., SOKAL E., VAN HOOFF F. Maturation of villus and crypt cell functions in rat small intestine. Role of dietary polyamines. *Dig. Dis. Sci.*, 1993, **38** : 1091-1098.
  13. DAHAN S., DALMASSO G., IMBERT V., PEYRON J., RAMPAL P., CZERUCKA D. *Saccharomyces boulardii* interferes with Enterohemorrhagic *Escherichia coli*-induced signaling pathways in T84 cells. *Infect. Immun.*, 2003, **71** : 766-773.
  14. MARANDI S., DE KEYSER N., STILMANT C., SALIEZ A., SOKAL E.M., GUIOT Y., BUTS J.P. Ontogeny of MAP kinases in rat small intestine : premature stimulation by insulin of BBM hydrolases is regulated by ERKs but not by p-38 MAP kinase. *Pediatr. Res.*, 2002, **52** : 180-188.
  15. KOLCH W. Coordinating ERK/MAPK signalling through scaffolds and inhibitors. *Nat. Rev. Mol. Cell Biol.*, 2005, **6** : 827-837.